A 63 year old woman visits her doctor with a three month history of fatigue and generalised joint pains. Her medical history is unremarkable and she reports no recent stress, infection, or weight loss. There are no abnormalities on clinical examination. Haemoglobin, creatinine, and electrolytes, liver enzymes, glucose, inflammatory markers, and thyroid function tests are normal. Ferritin, iron, transferrin, and transferrin saturation are also requested.

This article discusses some situations in which ferritin and iron studies might be helpful and how to avoid common pitfalls in their interpretation.

**What are the next investigations?**

The doctor in this case requested iron studies to investigate the possibility of iron overload and to screen for haemochromatosis. Iron studies are also commonly indicated in clinical practice to investigate iron deficiency, or to monitor response to treatment for these conditions (box 1).

Conventional laboratory tests of iron status are often referred to as "iron studies." They include tests for serum ferritin, iron, transferrin, or total iron binding capacity (TIBC), and transferrin saturation.

Iron is a key component of haemoglobin in red blood cells, myoglobin in muscle, and many metalloproteins and enzymes. It is essential for uptake of oxygen and its delivery to tissues, utilisation of oxygen by muscle cells, and mitochondrial energy production. Normal iron metabolism and regulation is outlined in Figure 1. The adult male body contains 3-5 g of iron. Less than 0.1% of total body iron stores circulate in plasma. Dietary Fe³⁺ is reduced to Fe²⁺ and transported into the enterocyte by the divalent metal transporter DMT1. Iron is exported across the basolateral membrane via the iron export protein ferroportin 1 or stored as ferritin. Transferrin bound iron binds to transferrin receptor 1 (TFR1) and is taken up into the cell via receptor mediated endocytosis. Expression of TFR1 is regulated locally by the iron demands of the cell, via binding of iron regulatory proteins. Old or damaged erythrocytes are removed from the circulation by splenic macrophages. Iron is removed from haem by haem oxygenase 1, and either stored as ferritin or released into the circulation.

Regulation of iron release at a systemic level is mediated by the peptide hormone hepcidin (produced predominantly by hepatocytes) and has an inhibitory effect on iron release from cells and dietary iron absorption. Expression is controlled by complex signalling pathways.

**What is included in iron studies?**

- **Ferritin**—is the intracellular storage form of iron. A very small amount is found in serum. In inflammation, liver disease, and malignancy, ferritin levels can rise because ferritin is an acute phase protein. In these patients, ferritin can appear either falsely high or normal, when in reality stores are low.
- **Serum iron**—refers to ferric ions (Fe³⁺) bound to serum transferrin. Serum iron concentration is highly variable and is affected by dietary iron intake, inflammation, and infection.
- **Transferrin**—is the principal iron transport protein in plasma. It increases in iron deficiency to maximise utilisation of available iron. Total iron binding capacity is an alternative test to transferrin. TIBC reflects the availability of iron binding sites on transferrin. Values increase in iron deficiency and decrease in iron overload. Some laboratories measure unsaturated iron binding capacity (UIBC) and calculate TIBC by adding serum iron to UIBC.
- **Transferrin saturation**—is calculated from serum iron and either TIBC or transferrin measurements. Typically, transferrin is 30% saturated with iron. Transferrin saturation rises in iron overload and falls in iron deficiency, but does not quantitatively reflect iron stores. A rise in serum iron due to dietary iron intake can cause raised transferrin saturation.
Interpretation

Interpretation of iron studies can be challenging because the difficulties listed above affect almost all markers of iron status. Nevertheless, iron studies play an important role in clinical assessment (table 1). Reference intervals are quoted; however, these can vary by laboratory and must be confirmed locally.

Iron overload

Testing for iron overload (increased total body iron stores with or without organ dysfunction) can be triggered by clinical features such as those listed in box 1. Primary iron overload includes inherited mutations in iron regulatory genes (causing iron loading syndromes such as haemochromatosis). Secondary iron overload is associated with other conditions or iatrogenic factors (box 2).

Hereditary haemochromatosis (an autosomal recessive genetic condition caused by mutation of the HFE gene) is the most common inherited cause of iron overload. Homozygosity for the C282Y polymorphism affects 1 in 200 people of northern European descent and accounts for more than 80% of clinically recognised cases. Clinical penetrance varies widely (1%-28% of C282Y homozygotes in population studies). In an unselected population, raised serum ferritin (>200 μg/L for premenopausal women or >300 μg/L for men and postmenopausal women) and transferrin saturation >50% diagnosed C282Y homozygosity with a sensitivity of 90% in men and 75% in women. An approach to hyperferritinaemia has previously been discussed. Specialist techniques for assessing iron overload include liver biopsy, magnetic resonance imaging, and superconducting quantum interference device susceptometry, a non-invasive technique that measures the amount of in vivo magnetisation caused by hepatic iron. Indications for these tests can include marked hyperferritinaemia (>1000 μg/L) or further assessment when the diagnosis of iron overload is in doubt. They are not widely available and are usually requested by specialists.
Investigating iron deficiency

Biochemical definitions are listed in box 3. Low serum ferritin (<15 μg/L) provides absolute evidence of iron deficiency. Iron deficiency can result from inadequate intake of iron, inadequate absorption or loss (through bleeding, whether frank or occult), or a combination of both. Prevalence varies by region and is often estimated using anaemia as an indirect indicator. It is quoted as 4.5%-18% in US based population studies. A fourth form of iron deficiency is functional. In such patients there are sufficient iron stores but they are inadequately utilised. Functional iron deficiency can occur in patients with acute and chronic infectious, inflammatory, or malignant conditions. Inflammatory cytokines increase the synthesis of hepcidin, which regulates iron. Hepcidin decreases iron absorption from the gastrointestinal tract and impedes mobilisation of iron from macrophages and hepatocytes. In certain patients—eg, those with chronic kidney disease, ferritin below the reference interval indicates deficient iron stores; however normal or high values cannot exclude functional iron deficiency. A retrospective study of more than 20 000 patient results found that serum ferritin rises, and transferrin and serum iron fall in patients with C reactive protein above 10 mg/L.

Diagnosis of functional iron deficiency in patients with an inflammatory condition can be useful in guiding further investigations and treatment (eg, iron supplementation), particularly when the patient has symptomatic anaemia or there is suspected occult blood loss. This is problematic, however, as there is no single reliable and widely available biochemical marker for functional iron deficiency. The British Committee for Standards in Haematology recommends percentage of hypochromic red cells (% HRC) as the best established variable for the diagnosis of functional iron deficiency. %HRC (>6%) is recommended for diagnosis of iron deficiency in chronic kidney disease if the sample can be processed within six hours; however, this might be precluded by availability of laboratory services. Reticulocyte haemoglobin content (<29 pg) is an alternative. Red cell zinc protoporphyrin, soluble transferrin receptor, and the soluble transferrin receptor log ferritin ratio have been proposed as markers of functional iron deficiency. They are not widely available, and soluble transferrin receptor assays are subject to difficulties with standardisation between laboratories. They are of limited use for the general clinician. Measurement of hepcidin as a diagnostic tool is primarily restricted to research at present; however, harmonisation of assays and validation in the clinical setting are ongoing.

Important points to consider when interpreting iron studies in the context of iron deficiency

- Suspected iron deficiency anaemia is best investigated using serum ferritin. Iron deficiency is confirmed by a level below the reference interval.

- Low serum iron cannot be interpreted in isolation because it might be seen in infection, inflammation, and malignancy as well as iron deficiency. We recommend measuring C reactive protein if serum ferritin is normal or high with serum iron or transferrin saturation below their respective reference intervals.

- Transferrin saturation <16% is poorly specific, as pregnancy, oral contraceptive use, and chronic illness can result in low transferrin saturation without iron deficiency.

- In general, avoid checking iron studies in those with acute inflammatory illness or when C reactive protein is known to be >10 mg/L. For patients with chronic inflammatory conditions, interpretation should be performed cautiously and results discussed with the specialist managing the condition or haematology where there is doubt.

Monitoring response to treatment

Serial measurements of serum ferritin are commonly requested to monitor iron status in patients who are receiving interventions to treat deficiency or prevent iron loading—eg, venesection and iron chelation. Which iron measure is chosen depends on the clinical situation.

Monitor ferritin in those with haemochromatosis undergoing venesection to deplete iron until ferritin is less than 50 μg/L, and maintain ferritin 50-100 μg/L thereafter. Iron studies might be requested to guide treatment with intravenous iron. In chronic kidney disease, serum ferritin concentration <100 μg/L in non-dialysed patients or <200 μg/L in chronic haemodialysis patients is associated with a high likelihood of iron deficiency and a potentially good response to intravenous iron treatment. Response to oral iron therapy in iron deficiency anaemia can usually be confirmed by monitoring the rise in haemoglobin, and re-checking iron studies is not routinely required.

Patient outcome

Results in this patient showed ferritin 682 μg/L (normal range 15-200), iron 34 nmol/L (10-30), transferrin 2.0 mg/L (2-3.5), and transferrin saturation 68% (25-45), in keeping with iron overload. After discussing genetic testing, the patient consented to HFE gene analysis. She was found to be homozygous for the C282Y mutation, confirming the diagnosis of hereditary haemochromatosis. She was referred to a specialist for further investigations and venesection therapy.

We have read and understood BMJ policy on declaration of interests and declare that we have no competing interests

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Patient consent not required.
Box 3: World Health Organization definitions of iron deficiency (2001)*

- Ferritin <15 μg/L
- OR transferrin saturation <16%
- OR haemoglobin increase of 1 g/dL after two months of iron supplementation (values vary with ethnicity and pregnancy)
- Anaemia is defined as haemoglobin <120 g/L in women and <130 g/L in men (15 years of age and above).

Education into practice

- How might your approach to testing for iron abnormalities in the presence of infection or inflammation change as a result of reading this article?
- Is serum ferritin used routinely in your place of work as an investigation for iron deficiency?
- When and how might you contact specialist teams for assistance with interpreting iron studies results?

How patients were involved in the creation of this article

- The idea for the article was conceived following a local survey of primary and secondary care clinicians, which highlighted that many clinicians feel uncomfortable when interpreting iron studies.
- Patients with iron related disorders were informally interviewed about their experiences with iron related blood tests during investigation and diagnosis. Some expressed the view that they would have valued more detailed explanation of their test results, which helped inspire the content of this article.

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# Table

Table 1: Serum measures of iron status in states of deficiency and overload

<table>
<thead>
<tr>
<th></th>
<th>Serum ferritin</th>
<th>Serum iron</th>
<th>Transferrin/total iron binding capacity (TIBC)</th>
<th>Unsaturated iron binding capacity</th>
<th>% transferrin saturation</th>
</tr>
</thead>
<tbody>
<tr>
<td>Normal values</td>
<td>15-200 μg/L (premenopausal women)</td>
<td>10-30 nmol/L</td>
<td>Transferrin 2.0-3.5 g/L TIBC 45-81 μmol/L</td>
<td>12-43 μmol/L (men)</td>
<td>13-56 μmol/L (women)</td>
</tr>
<tr>
<td>Haemochromatosis with iron overload</td>
<td>↑</td>
<td>↑</td>
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<tr>
<td>Secondary iron overload (eg, repeated transfusion)</td>
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<td>↑</td>
<td>↓/↔</td>
<td>↓/↔</td>
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<tr>
<td>Iron poisoning</td>
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<tr>
<td>Iron deficiency</td>
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<td>↔/↑</td>
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<td>↓</td>
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<tr>
<td>Anaemia of chronic disease/inflammatory response</td>
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Figure

**Fig 1** Overview of iron absorption, transport, utilisation, and excretion.